

Research Article

HISTOPATHOLOGICAL STUDY OF DIFFERENT ORGANS OF CHARLES FOSTER RATS UNDER THE EXPOSURE OF *PUERARIA TUBEROSA*

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ABSTRACT: The present study was undertaken to investigate the safe doses of *Pueraria tuberosa* water extract (PTWE) on different organs. The OECD guidelines 407 of repeated toxicity was followed with respect to the selection of dose and days for different organs. The selected doses of PTWE were 250, 500, 1000 and 2000 mg/kg b wt for 7, 14, 21 and 28 days. Haematoxylin and eosin staining was used to study the morphological alterations in heart, intestine, testis, adrenal gland and spleen. In the present study, no adverse alterations in cardiac fibers of the heart, size and shapes in crypts and villi of intestine, seminiferous tubules and spermatozoa count in testis, three zones of adrenal gland, and spleen were seen in all treated groups of PTWE. There were no adverse morphological alterations found in described organs. The PTWE are safe at 1000 mg/kg b wt. up to 28 days and 2000 mg/ kg b. wt up to 21 days, respectively.

Key words: Histopathological alteration, OECD, Organ toxicity, *Pueraria tuberosa*, Water extract, Charles Foster rats.

INTRODUCTION

In several countries, more than 60% of the population are dependent on herbal medicine for health care (Saminathan *et al.* 2013). However, because of non-availability of safety parameters, many people are hesitant in using herbal medicine. Although it is well established that herbal medicines have therapeutic responses and they are already in clinical use (Saminathan *et al.* 2013).

The powder of *Pueraria tuberosa* (PT) tubers is in clinical use in Ayurveda as health promotion medicine (Sherman *et al.* 2010). In Hindi, it is called as vidarikand and in English it is known as Indian kudzu. It belongs to the family Fabaceae (Prasain *et al.* 2012). Its major secondary metabolites include puerarin, tuberostan, genestein, daidzin, tuberosin, puerarin, and pterocarpan puerarone (Rastogi *et al.* 2013, Maji *et al.* 2014a,b). In kudzu root, puerarin is the most abundant (approx. 23% w/w) and it has potent ability to cause various pharmacological effects (Lee *et al.* 2005). *Pueraria tuberosa* has shown many pharmacological activities in rats (Pandey *et al.* 2020). Pharmacokinetic study of PT was studied by Pandey *et al.* (2019b).

Recent past, it has been shown that *Pueraria tuberosa* water extract (PTWE) act as antidiabetic herbal drug working through incretin signalling pathway (Srivastava *et al.* 2018). Further, it was reported regarding its mechanism of action towards anti-diabetic and nephroprotective potential by inhibition of DPP4, MMP-9, PKC-beta, and activation of Caspase enzymes (Tripathi *et al.* 2016). The results have shown activation of SOD, catalase, BCL-2, and nephrin (Srivastava *et al.* 2015, Shukla *et al.* 2016, Srivastava *et al.* 2017). However, for the first time we reported its action of GLP-1, GIP, TNF-alpha, IL-6 and HIF-1 which were inhibited by *Pueraria tuberosa* in different experimental conditions such as diabetes and kidney damage (Srivastava *et al.* 2019). These results were according to earlier reports related to the role of different pure phytochemicals, which are also found in PT tubers (Bebrevska *et al.* 2007, Cai *et al.* 2011).

The histopathological examination is the golden standard for evaluating treatment related pathological changes in tissues and organs (OECD 1995). The pathological study can add information to the clinical data. Histopathological study is very clear, reliable

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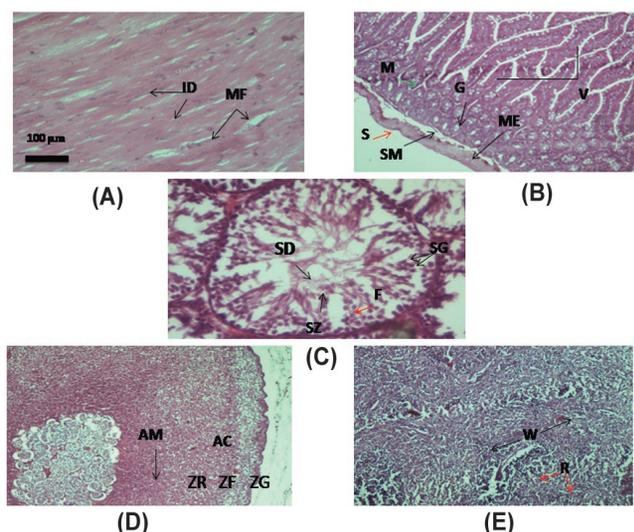


Fig. 1. Histology of normal rats (A) Heart, (B) Intestine, (C) Testis, (D) Adrenal gland and (E) Spleen.

[Scale bar 100 μ m with different magnification like 10x and 40X according to specific organs site .ID-Intercalated disc, MF-Myofibrils, G-Glands, M-Mucosa, SM-Sub mucosa, S-Serosa, ME-Muscularia externa, V-Villi, SG-Spermatogonia, SD-Spermatids, AM-Adrenal medulla, AC- Adrenal cortex, ZR-Zona glomerulosa, ZF-Zona fasciculata, ZG- Zona reticularis, R-Red pulp, W-white pulp].

parameter to study the effectiveness/toxicity of drugs in animal tissues. Study of safety and toxicity are the main parameters used before the clinical use of drugs.

Earlier, we reported the effect of PTWE in pre-clinical toxicity in rats as per OECD guideline 425 and 407 with special reference to changes in liver and kidney functions (Brondani *et al.* 2017, Pandey *et al.* 2018a). In the present study, partially purified water extract of tubers of *Pueraria tuberosa* (PTWE) has been used to assess its toxic effect on Charles Foster rats as per OECD guidelines. The herbal tablets of PTWE were prepared by a wet granulation method for clinical study (Pandey *et al.* 2018b). In the present study, effect of *Pueraria tuberosa* on other organs after oral treatment with different doses for different durations up to 28 days. The histological changes have been observed in heart, intestine, testis, adrenal gland and spleen in rats of Charles foster strain.

MATERIALS AND METHODS

Collection of plant material and preparation of extract

The plant material *Pueraria tuberosa* (PT) was purchased from Ayurvedic Pharmacy BHU Varanasi, preserved in our laboratory, Department of Medicinal

Chemistry IMS BHU (Ref. no YBT/MC/12/1-2007) and also with the sample preserved in Museum of Department of Dravya-guna of our Institute IMS BHU. It was further compared with Thin Layer Chromatography finger printing with standard sample, which has been characterized earlier in our laboratory Department of Medicinal Chemistry by DNA fingerprinting, High Performance Thin Layer Chromatography and Light Chromatography-Mass Spectrometry (Nagwani *et al.* 2010). Pharmacognostic characterization of crude powder of PT was done on the basis of standard pharmacopeia (Pandey *et al.* 2019a). The water extract of plant *Pueraria tuberosa* was prepared by water decoction method. The yield value of extract was 36% w/w. It was characterized by TLC finger printing.

Animals

Charles Foster strain of rats were purchased from the Central Animal House of our Institute IMS BHU (542/GO/ReBi/S/02/CPCSEA). The animals were acclimatized for 7 days in laboratory condition and subjected to anti-protozoa treatment by giving drug Metronidazole orally. Finally, the animals were randomly divided into five groups of 6 animals in each. The experimental protocol was approved by the Institutional Ethical Committee (Dean/2017/CAEC/721).

Experimental design

The selection of dose and time of drug treatment was followed as per earlier studies and the experiment was carried out as per OECD guidelines 407 (Brondani *et al.* 2017). The dose was prepared by dissolving PTWE in water with gum acacia and different dose (250 mg/kg b. wt, 500 mg/kg b. wt, 1000 mg/kg b. wt, and 2000 mg/kg b. wt) were orally given for 28 days, to each rat of the respective groups in the morning time. Weekly assessments of body weight and diet intake were recorded. At the end, the animals were sacrificed by anesthetizing the rats by intra-peritoneal injection of pentobarbitone sodium (45 mg/kg b. wt). The blood was collected immediately after death, and all the required organs were dissected out. The attached fat and other undesired tissues were dissected out and finally the organs were drained on the blotting paper and weighed on electronic pan balance. The chosen organs were heart, intestine, testis, adrenal gland and spleen, which were finally fixed in formalin for histopathological study. The animals of different groups were sacrificed at different time intervals of 7, 14, 21 and 28 days. The tissues of the group having a dose of 2000 mg/kg b. wt could not be collected at 28th day, because all animals of this group died.

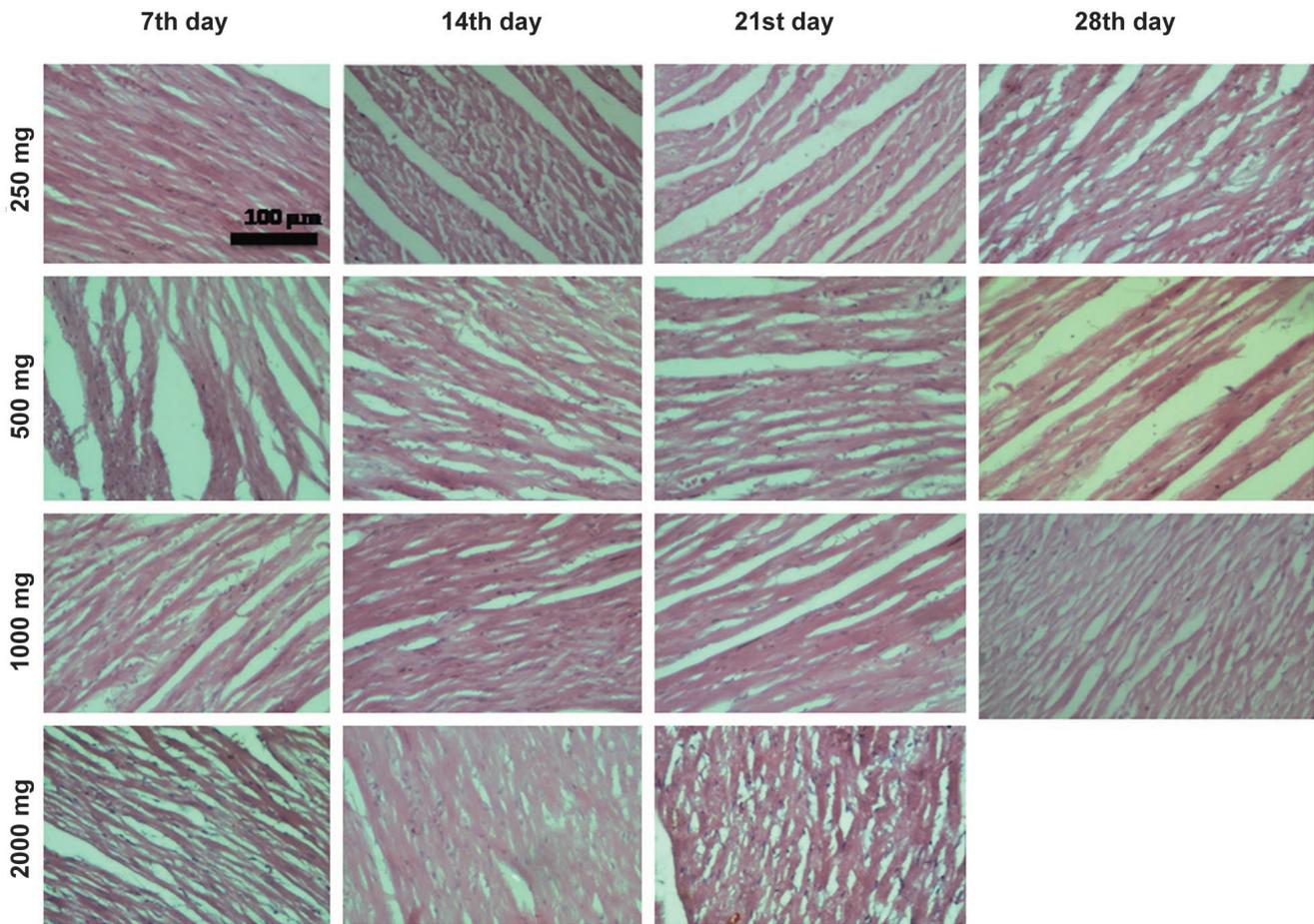


Fig. 2. Histology of PTWE treated heart tissue at different time intervals with variable dosage. Scale bar 100 μ m.

Histopathology

Organs such as heart, spleen, testis, intestine, and adrenal gland were isolated and trimmed of excess fat in each group of animals, fixed in formalin for H & E staining. The standard protocol of making micro sections and staining was adopted (Troyer 2008). The fixed tissues were taken out of the fixative, washed properly and small sections were processed for dehydration and finally embedded in the paraffin wax. After solidification, the blocks were trimmed, mounted on microtome (medimeas /mrm-1120 A) and thin sections of 5-6 μ m thickness were cut and placed on slides coated with albumin and then finally subjected to a process of dewaxing, dehydration and staining with haematoxylin and eosin (H &E) staining. Finally, the sections were mounted under a cover slip and sealed with DPX Mount. The transverse sections of all the organs were examined and image were taken with binocular fluorescent microscope without lamp on (Nikon Eclipse 50i Japan), fitted with a digital camera. Histological analysis was done to further confirm the alteration in cell structure of the organs. All the slides

were randomly photographed in 10 view fields. The measurements of captured photographs were done by using software NIS Elements Basic Research.

RESULTS AND DISCUSSION

The gross lesions in heart, intestine, testis, adrenal gland and spleen were found to be normal as earlier reported. Most of them showed a high degree of liver toxicity, as already reported in our preclinical toxicity study (Pandey *et al.* 2018a).

Control animals

The heart tissue section had normal intact cardiac muscle myofibrils and intercalated disc (Fig. 1A). In the intestinal tissue, glands, mucosa, sub-mucosa, serosa, muscularis externa and villi were normal (Fig. 1B). The spermatogonia, spermatozoa, and spermatids were normal in testis (Fig. 1C). Adrenal gland showed normal outermost portion, cortex and inner section, medulla (Fig 1D). Splenic section showed normal red and white pulp areas (Fig. 1E).

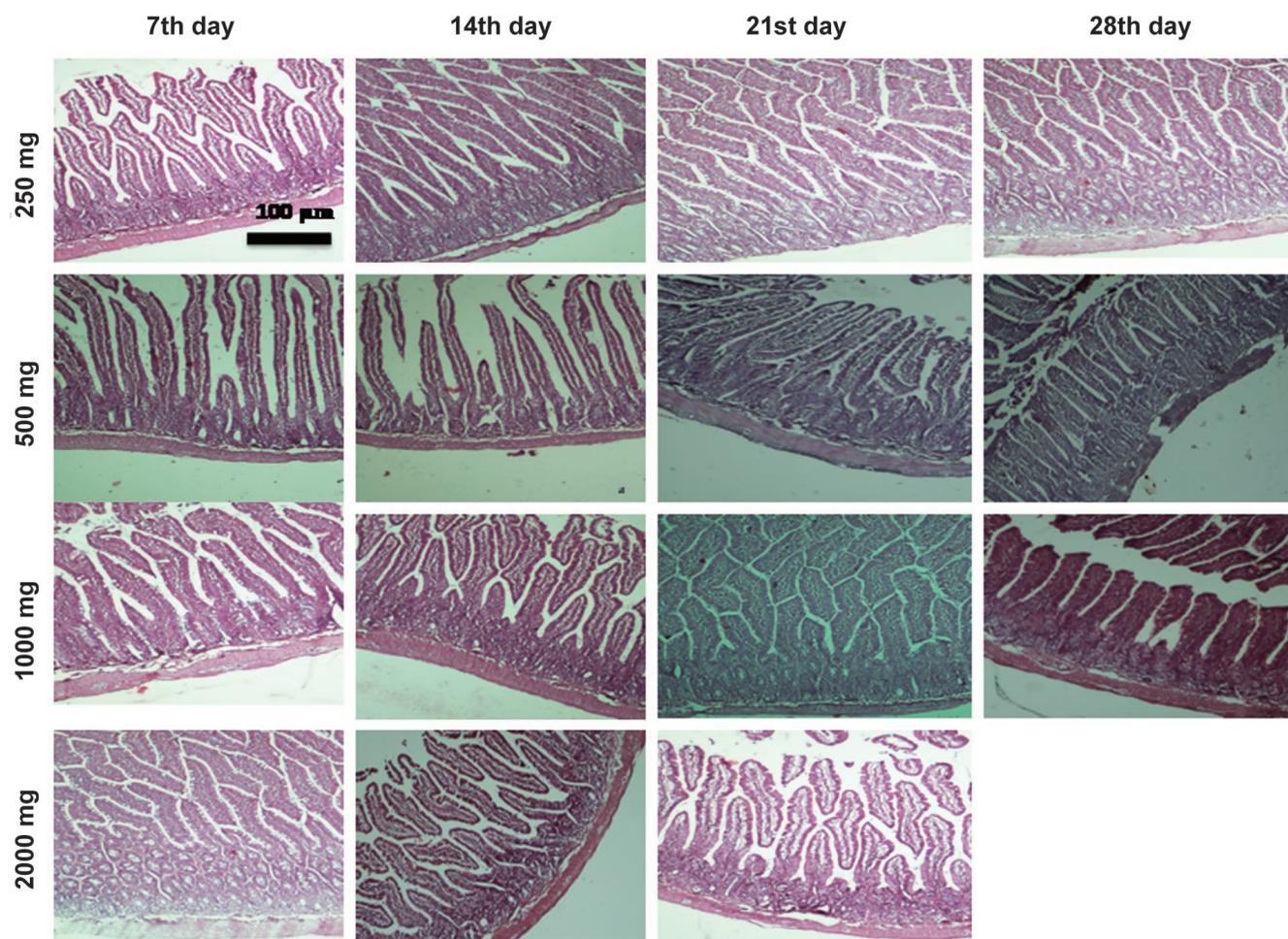


Fig. 3. Histology of PTWE treated intestinal tissue at different time intervals with variable dosage. Scale bar 100 μ m.

Effects of PTWE on different organs at different doses

Heart The T.S. of heart did not show any changes in the extract treated group for all tested doses up to 21 days. The sections showed intact cardiac muscle myofibrils and intercalated disc (Fig. 1A). The treatment for 21 days with 2000mg/kg b wt also did not show any cellular damage or focal necrosis of cardiac muscle fibres. No interstitial oedema or swelling was also found (Fig. 2)

Intestine (Jejunum)

The intestinal part, which was selected for study was 10 cm distal to the duodenum. Jejunum showed no significant changes in the PTWE treated rats. The T.S. images were similar to normal rat intestine (jejunum) The serosa and mucous membranes were normal. The number and size of villi were also normal. The glands found in the intestinal wall were also normal (Fig. 3).

Testis

Histopathological examination of the testis showed

normal histological structure of active mature, functioning seminiferous tubules associated with complete spermatogenic series are shown in Fig.1 and Fig.4. The spermatozoa and spermatogonia were found in fully mature condition in all sections of tissue of doses up to 2000mg/kg b wt for 21 days. There were no congestion, edema and dilation were found in blood vessels (Fig.4). It may be effective in patients, already having a defective physiology.

Adrenal gland

The zona glomerulosa, zona fasciculata, zona reticularis and adrenal medulla were intact in all tissue sections. No morphological deviation was found in the cortex region of the adrenal gland. There was no amorphous debris found in focal areas of zona fasciculata of the higher dose of PTWE *i.e.*, for 100 and 2000mg/kg bw up to 21 and 28 days respectively (Fig.5).

Spleen

The spleen showed the normal histo-architecture in red pulp and white pulp regions in all the tested doses of

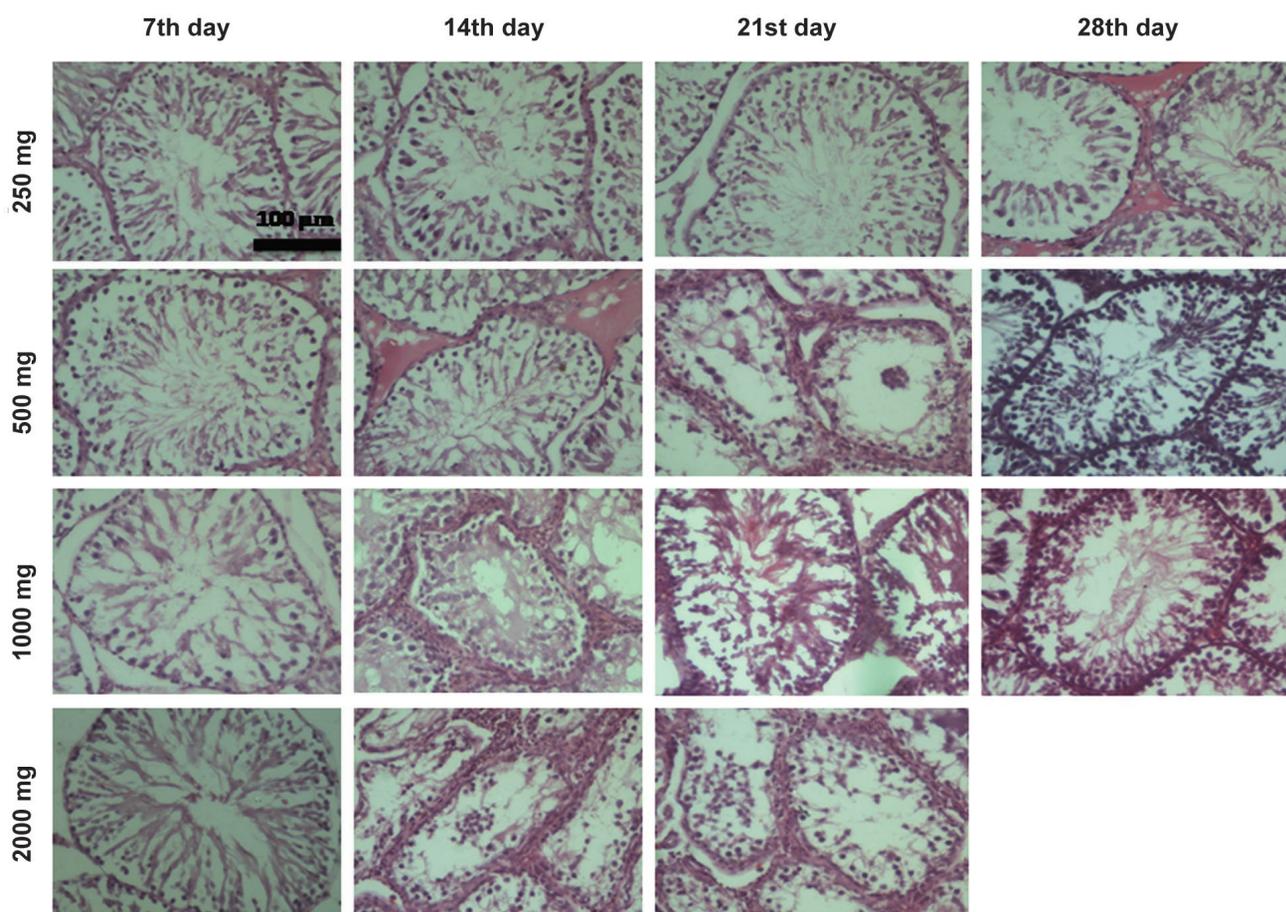


Fig. 4. Histology of PTWE treated testis tissue at different time intervals with variable dosage. Scale bar 100 µm.

PTWE. The red pulp region consisted of red blood cells, lymphocytes and plasma cells, whereas the white pulp region contained T lymphocytes and B lymphocytes. (Fig.6).

As compared to growing the number of herbal drug users around the globe, there is a lack of scientific data on the safety profile of herbal products (Saad *et al.* 2006), therefore the safety of these products have become an important issue (WHO 2004). In this experiment, we have conducted OECD guidelines 407 and found that histopathological examination of the vital organs did not reveal any morphological changes after oral administration of *Pueraria tuberosa* for 28 days at the dose level of 250, 500, 1000 and 2000 mg/kg bw.

In the present scenario, cardiovascular diseases, particularly, become a worldwide health problem affecting all economic groups of the society.

Tannins, flavonoids and glycosides have significant antioxidant properties, thus augments antioxidants and induction of HSP 72 (Nieto *et al.* 1993). In the previously reported isoflavonoids and glycosides are found in plant *Pueraria tuberosa*. PT is also reportedly known for its

antioxidant property (Pandey *et al.* 2007). The major bioactive component of the PT is flavonoids known as puerarin. On the basis of previous study of puerarin and our present result, we can assume puerarin might be the possible component of the PT which protect the cardiovascular tissue via a different mechanism as puerarin lower the mRNA and protein level and its receptor APJ in one clip hypertension (Jin *et al.* 2009), and blocks TSP 1 expression in diabetic rats (Pan *et al.* 2009).

The Intestine is the primary organ responsible for food materials and drugs absorptions. The small intestine comprises 4 layers, mucosa, sub mucosa, muscularis externa and serosa. Mucosa consists of 3 layers, epithelium, lamina propria and muscularis mucosae organized into villi and crypts (crypts of Lieberkuhn). Villi were finger like projections of the epithelium which contained blood and lymphatic vessels. Intestinal mucositis is a common side effect of clinical chemotherapy for patients with cancer (Wadler *et al.* 1998), and includes symptoms such as severe diarrhoea and dehydration. The anti metabolite anticancer agent,

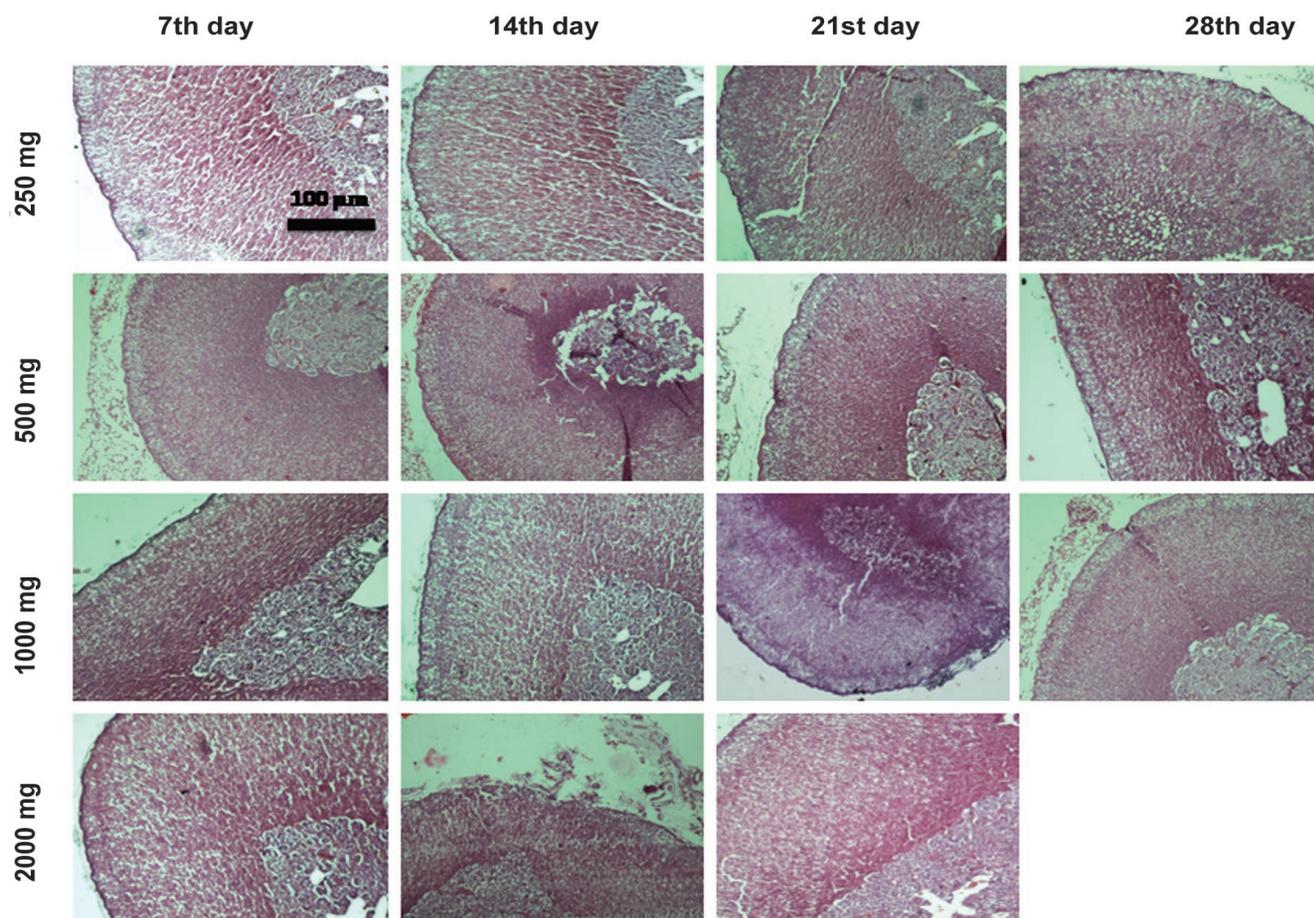


Fig. 5. Histology of PTWE treated adrenal tissue at different time intervals with variable dosages. Scale bar 100 μ m.

5-fluorouracil (5-FU), is widely used to treat several types of malignant tumours and it frequently causes intestinal mucositis. Mucositis is morphologically characterized by the shortening of villus height and destruction of crypts in the small intestine. Previously the apoptosis was detected in intestinal crypts 24 hours after the first administration of 5-FU in mice. Those puerarin responsible for anti-inflammatory activities via inhibiting the level of IL-8 (Pang *et al.*, 2012) suppressed the protein or the mRNA expression of TNF- α , NF- κ B, iNOS, TGF- β 1 and MDA (Li *et al.* 2013). In our result, no significant adverse effect was found in different doses of *Pueraria tuberosa* on morphological change in size and shapes of the villi and crypts of intestine since it was reported that PT have more isoflavonoids.

The study results related with testis were contrary to the literature, showing its anti-infertility and spermatogenic activating potential, at least in normal rats. It may be effective in patients, already having a defective physiology.

Drug induced toxicity was found in the testis by cisplatin and metabolic disorder such high fat diet induced diabetic model in rats. Various types of isoflavones,

phyto-estrogens, puerarin etc. responsible for the stimulation of androgenic activity have been reported in the roots of *Pueraria tuberosa* (Chauhan *et al.* 2011, Chauhan *et al.* 2013). Phytoestrogen like daidzein and genistein also affecting neurobehavioral aspect are anti-oestrogenic with an action opposite to that of oestradiol and increase the level of LH, FSH, and testosterone via stimulation of gonadotrophin releasing hormones GnRH (Lee *et al.* 2002, Patisaul 2005). Kudzu and puerarin combination with p4 may synergistically interfere with NADPH oxidation reported in chromatin condensation (Chapman and Michael 2003, Bennetts *et al.* 2008). In our results any types of morphological changes were not found in rats after 28 days treatment of PTWE.

We know that in adrenal gland necrosis found more frequently in the cortex (especially zona fasciculata and reticularis) than medulla. Flavonoids like Daidzein and nobiletin those found in PT was earlier reported for less, but significant increase in catecholamine synthesis or secretion via activation of extracellular signal regulated protein kinases (ERKs) through the plasma membrane estrogen receptor (Liu *et al.* 2007), thus enhance the symaptho-adrenal system. The histological changes in

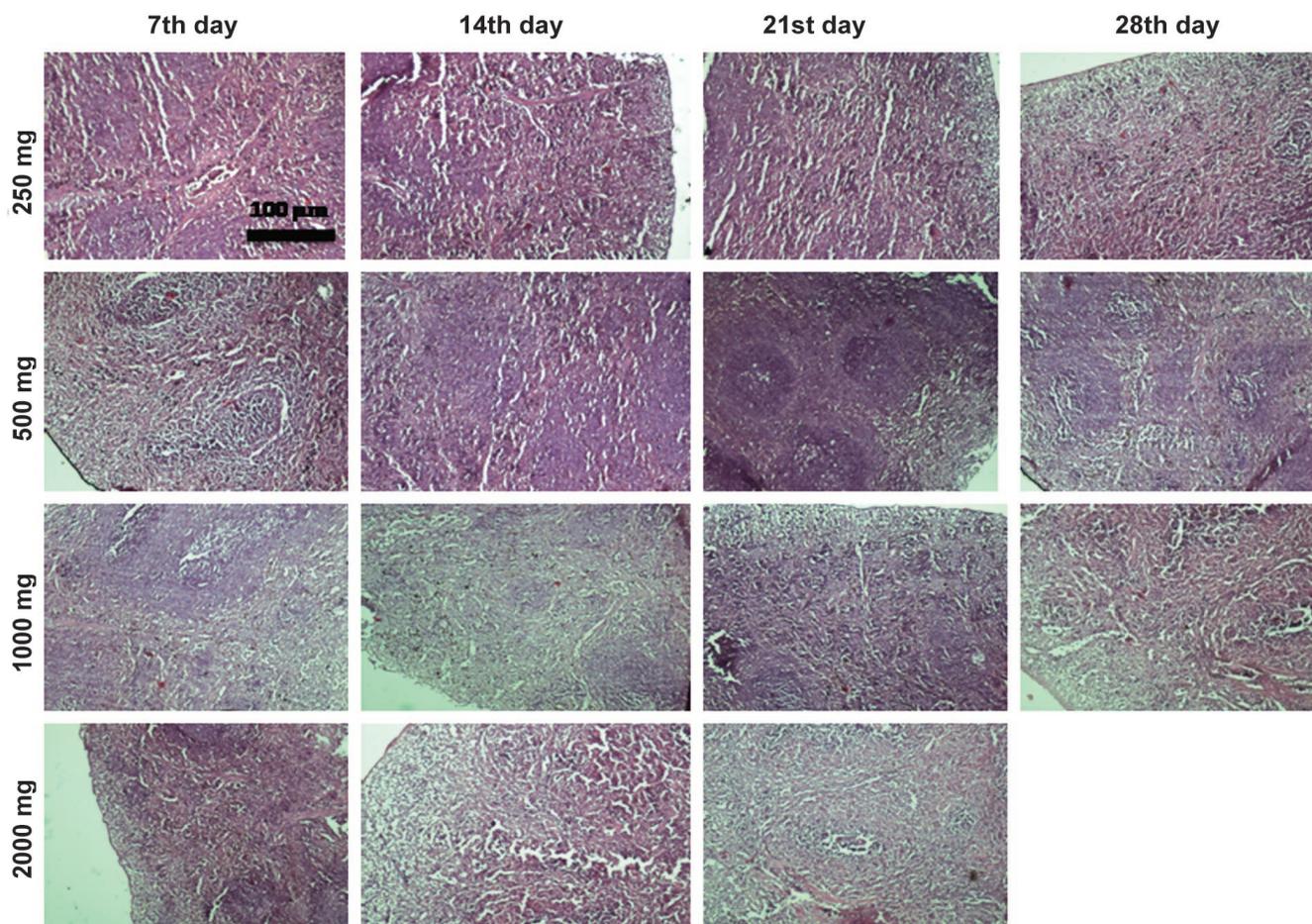


Fig. 6. Histology of PTWE treated splenic tissue at different time intervals with variable dosages. Scale bar 100 μ m.

adrenal could be correlated to its function. It is already reported that the release of the corticosteroids cortisol and aldosterone can be stimulated through the sympatho-adrenal system, by mediation through chromaffin cells in a paracrine manner. But here no such changes have been observed in our study, suggesting no adverse effect of PTWE on these systems.

The isoflavones like puerarin, daidzein, and genistein are the most important constituents, and these are responsible for the immunomodulatory function (Sawale *et al.* 2013). The effect of *Pueraria tuberosa* and its isoflavones on haematopoietic system as well as on the function of T cells and neutrophils have been extensively correlated with immunomodulatory function.

The leukocytes including neutrophils, lymphocytes, monocytes, eosinophils, and basophils are responsible for immune response. In our experimental study histopathological investigation of spleen did not exhibit any abnormalities treated with a low or high dose of Pueraria extract. Also, spleen appeared grossly coloured and no congestion was found in all treated tissues. The tissue

was clearly differentiated into white pulp (WP) and the red pulp (RP). The architecture of the white pulp displayed normal rounded scattered follicles.

CONCLUSION

No alterations were founded in cardiac fibers in the heart, size and shapes in crypts of intestine, seminiferous tubules and spermatozoa were normal in testis. Three zones of adrenal gland were normal, and no adverse changes were seen in the pulps of spleen in all the groups treated with 2000 mg/kg b. wt of PTWE up to 21 days and with 1000 mg/kg b. wt of PTWE up to 28 days. The experiments were found to be safe up to 2000mg/kg b. wt. treatment of PTWE after 21 days.

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REFERENCES

- Bebrevska L, Bravo L, Vandervoort J, Pieters L, Vlietinck A, Apers S (2007) Development and validation of an HPLC method for quality control of *Pueraria lobata* flower. *Plant Medicine* 73(15): 1606-1613.
- Bennetts LE, De Iuliis, GN Nixon B, Kime M *et al.* (2008) Impact of estrogenic compounds on DNA integrity in human spermatozoa: Evidence for cross-linking and redox cycling activities. *Mutat Res Fundam Mol Mech Mutagen* 641: 1-11.
- Brunnicardi F, Andersen D, Billiar T *et al.* (2010) Schwartz's Principles of Surgery, 9th edn. 978.
- Cai RL, Li M, Xie SH, Song Y, Zou Z *et al.* (2011) Antihypertensive effect of total flavone extracts from *Puerariae Radix*. *J Ethnopharmacol* 133: 177-183.
- Brondani JC, Reginato FZ, da Silva Brum E, de Souza Vencato M, Lhamas CL *et al.* (2017) Evaluation of acute and subacute toxicity of hydroethanolic extract of *Dolichandra unguis-cati* L. leaves in rats. *J Ethnopharmacol* 202: 147-153.
- Chapman JC, Michael SD (2003) Proposed mechanism for sperm chromatin condensation/decondensation in the male rat. *Reprod Biol Endocrinol* 1(20): 01-07.
- Chauhan NS, Gupta NK, Sharma V, Dixit VK (2011) Spectrofluorimetric estimation of puerarin in *Pueraria tuberosa*. *Acta Pol Pharm - Drug Res* 68: 453-456.
- Chauhan NS, Sharma V, Thakur M, Dixit VK *et al.* (2013) *Pueraria tuberosa* DC extract improves androgenesis and sexual behavior via FSH LH cascade. *Sci World J* 780659: 8.
- Gao Q, Yang B, YZ Guo, Wang J, Bruce LC *et al.* (2007) Opening the calcium-activated potassium channel participates in the cardioprotective effect of puerarin. *Eur J Pharmacol* 574: 179-184.
- Gray SL, Lackey BR, Boone WR (2015) Impact of kudzu and puerarin on sperm function. *Reprod Toxicol* 53: 54-62.
- Jin G, Yang P, Gong Y, Fan X, Tang J *et al.* (2009) Effects of puerarin on expression of apelin and its receptor of 2K1C renal hypertension rats. *Zhongguo Zhong Yao Za Zhi* 34: 3263-3267.
- Lee HU, Bae E A, Kim DH (2005) Hepatoprotective effect of Tectoridin and Tectorigenin on tert-Butyl Hydroperoxide-induced liver injury. *J Pharmacol Sci* 97: 541-544.
- Lee JS., Mamo J, Ho N, Pal S (2002) The effect of *Puerariae radix* on lipoprotein metabolism in liver and intestinal cells. *BMC Complement Altern Med* 2:12.
- Li R, Xu L, Liang T, Li Y, Zhang S (2013) Puerarin mediates hepatoprotection against CCl₄-induced hepatic fibrosis rats through attenuation of inflammation response and amelioration of metabolic function. *Food Chem Toxicol* 52: 69-75.
- Liu M, Yanagihara N, Toyohira Y, Tsutsui M, Ueno S (2007) Dual effects of daidzein, a soy isoflavone, on catecholamine synthesis and secretion in cultured bovine adrenal medullary cells. *Endocrinology* 148: 5348-5354.
- Maji AK, Mahapatra S, Banerjee D (2014a) *In-vivo* immunomodulatory potential of standardized *pueraria tuberosa* extract and its isoflavonoids. *Int J Pharm Sci* 6: 861-867.
- Maji AK, Pandit S, Banerji P, Banerjee D (2014b) *Pueraria tuberosa*: A review on its phytochemical and therapeutic potential. *Nat Prod Res* 23: 2111-2127.
- Nagwani S, Tripathi YB (2010) Amelioration of cisplatin induced nephrotoxicity by PTY: A herbal preparation. *Food Chem Toxicol* 48: 2253-2258.
- Ng CF, Koon CM, Cheung DWS, Lam MY, Leung PC *et al.* (2011) The anti-hypertensive effect of Danshen (*Salvia miltiorrhiza*) and Gegen (*Pueraria lobata*) formula in rats and its underlying mechanisms of vasorelaxation. *J Ethnopharmacol* 137: 1366-1372.
- Nieto S, Garrido A, Sanhueza J, Loyola LA, Morales G *et al.* (1993) Flavonoids as stabilizers of fish oil: An alternative to synthetic antioxidants. *J Am Oil Chem Soc* 70: 773-778.
- Organisation for Economic Co-operation and Development (1995) Recommendation of the Council of the OECD on Improving the Quality of Government Regulation (Adopted on 9 March 1995). *OECD* 55: 1016-1031.
- Pan ZY, Bao ZS, Wu ZM, Wang XM, Zheng JZ *et al.* (2009) The myocardial protective effects of puerarin on STZ-induced diabetic rats. *Mol cell Biol* 42: 137-144.
- Pandey H, Srivastava S, Kumar R, Tripathi YB (2018a) Preclinical acute and repeated dose toxicity of *Pueraria tuberosa* (PTWE) on charles foster rats. *Int J Pharm Sci Res* 9: 4572-4581.
- Pandey H, Srivastava S, Mishra B, Saxena R, Tripathi YB (2018b) Development and evaluation of herbal tablet loaded with *Pueraria tuberosa* water extract with use of different Excipients. *Asian J Pharm* 12: 786-793.

- Pandey H, Srivastava S, Dwivedi M, Upadhyay N, Singh M (2019) Pharmacognostic standardization of tuber *Pueraria tuberosa*. J Imer Tech Innov Res 6(4): 611-622.
- Pandey H, Srivastava S, Kumar A, Kumar R, Tripathi YB (2019) Pharmacokinetics study of Puerarin absorption in blood after consumption of *Pueraria tuberosa* water extract (PTWE) by rats. Explor Anim Med Res 9(2): 188-196.
- Pandey H, Srivastava S, Tripathi YB (2020) A novel approaches for drug development and pharmacological study of herbal plant. Int J Pharm Sci Res 11(12): 5974-5986.
- Pandey N, Chaurasia JK, Tiwari OP, Tripathi YB (2007) Antioxidant properties of different fractions of tubers from *Pueraria tuberosa* Linn. Food Chem. 105: 219–222.
- Pang W, Lan XM, Wang CB (2012) Effect of puerarin on the release of interleukin-8 in co-culture of human bronchial epithelial cells and neutrophils. Chin J Integr Med 18: 283-287.
- Patisaul HB (2005) Phytoestrogen action in the adult and developing brain. J Neuroendocrinol 17: 57-64.
- Prasain JK, Peng N, Rajbhandari R, Michael Wyss J (2012) The Chinese Pueraria root extract (*Pueraria lobata*) ameliorates impaired glucose and lipid metabolism in obese mice. Phytomedicine 20: 17-23.
- Rani VU, Sudhakar M, Ramesh A (2017) Protective effect of *Pueraria tuberosa* Linn. in arsenic induced nephrotoxicity in rats. Asian J Pharm Res 7: 15.
- Rastogi S, Katara A, Pandey MM, Arora S, Singh RRB (2013) Physical stability and HPLC analysis of Indian Kudzu (*Pueraria tuberosa* L.) fortified milk. Evidence-based Complement Altern Med 2013: 368248.
- Saad B, Azaizeh H, Abu-Hijleh G, Said O (2006) Safety of traditional Arab herbal medicine. Evidence-Based Complement. Altern Med 3: 433-439.
- Saminathan M, Rai RB, Dhama K, Tiwari R, Chakraborty S *et al.* (2013) Systematic review on anticancer potential and other health beneficial pharmacological activities of novel medicinal plant *Morinda citrifolia* (Noni). Int J Pharmacol 9(8): 462-492.
- Sawale PD, Singh RRB, Kapila S, Arora S, Rastogi S *et al.* (2013) Immunomodulatory and antioxidative potential of herb (*Pueraria tuberosa*) in mice using milk as the carrier. Int J Dairy Technol 66: 202-206.
- Sherman PW, Billing J, Pande GS, Chuneekar KC, Nunn N *et al.* (2010) Bhavprakash Nighantu Flavour 23: 634.
- Srivastava S, Koley TK, Singh SK, Tripathi YB (2015) The tuber extract of pueraria tuberosa Linn. competitively inhibits DPP-IV activity in normoglycemic rats. Int J Pharm Sci 7: 227-231.
- Srivastava S, Shree P, Tripathi YB (2017) Active phytochemicals of *Pueraria tuberosa* for DPP-IV inhibition: *in silico* and experimental approach. J Diabetes Metab Disord 16(1): 46.
- Srivastava S, Shree P, Pandey H, Tripathi YB (2018) Incretin hormones receptor signaling plays the key role in antidiabetic potential of PTY-2 against STZ-induced pancreatitis. Biomed Pharmacother 97: 330-338.
- Srivastava S, Pandey H, Singh S K, Tripathi YB (2019) Anti-oxidant, anti-apoptotic, anti-hypoxic and anti-inflammatory conditions induced by PTY-2 against STZ-induced stress in islets. Bioscience Trends. 13 (5): 382-393.
- Tripathi YB, Shukla R, Pandey N, Pandey V, Kumar M (2016) An extract of *Pueraria tuberosa* tubers attenuates diabetic nephropathy by upregulating matrix metalloproteinase-9 expression in the kidney of diabetic rats. J Diabetes 9: 123-132.
- Troyer D (2008) Biorepository standards and protocols for collecting, processing, and storing human tissues. Methods Mol Biol 441: 193-220.
- Tsuchihashi R, Koder M, Sakamoto S, Nakajima Y, Yamazaki T *et al.* (2009) Microbial transformation and bioactivation of isoflavones from Pueraria flowers by human intestinal bacterial strains. J Nat Med 63: 254-260.
- Wadler S, Benson AB, Engelking C, Catalano R, Field M *et al.* (1998) Recommended guidelines for the treatment of chemotherapy-induced diarrhea. J Clin Oncol 16(9): 3169-3178.
- Wong KH, Li GQ, Li KM, Razmovski-Naumovski V, Chan K (2011) Kudzu root: traditional uses and potential medicinal benefits in diabetes and cardiovascular diseases. J Ethnopharmacol 134(3): 584-607.
- World Health Organization (2004) WHO guidelines on safety monitoring of herbal medicines in pharmacovigilance systems, Pharmacovigil Syst 17–20.

Xia DZ, Zhang PH, Fu Y, Yu WF, Ju MT (2013) Hepatoprotective activity of puerarin against carbon tetrachloride-induced injuries in rats: a randomized controlled trial. *Food Chem Toxicol* 59: 90–95.

Yan LP, Chan SW, Chan ASC, Chen SL, Ma XJ *et al.* (2006) Puerarin decreases serum total cholesterol and enhances thoracic aorta endothelial nitric oxide synthase expression in diet-induced hypercholesterolemic rats. *Life Sci* 79: 324-330.

Zhang H, Zhang L, Zhang Q, Yang XC, Yu JY *et al.* (2011) Puerarin: A novel antagonist to inward rectifier potassium channel (I K1). *Mol Cell Biochem* 352: 117-123.

Zhang J, Li X, Gao Y, Guo G, Xu C *et al.* (2013) Effects of puerarin on the inflammatory role of burn-related procedural pain mediated by P2X7 receptors. *Burns* 39: 610-618.

Zhang R, Hu Y, Yuan J, Wu D (2009) Effects of *Puerariae radix* extract on the increasing intestinal permeability in rat with alcohol-induced liver injury. *J Ethnopharmacol* 126: 207-214.

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